

Basic Immunology

25th lecture:

Immunology of periodontal diseases

Periodontal diseases

Diseases affecting the gingiva and supporting structures of teeth

Results in attachment loss and destruction of alveolar bone

Etiology is important for proper treatment



Marginal gingivitis



Classification of periodontal diseases (AAP, 1999)

I. Gingival diseases

- A. Plaque induced

- B. Non-plaque induced

II. Chronic periodontitis

- A. Localized

- B. Generalized

III. Aggressive periodontitis

- A. Localized

- B. Generalized

IV. Periodontitis as a manifestation of systemic disease

V. Necrotizing periodontal diseases

VI. Abscesses of the periodontium

VII. Periodontitis associated with endodontic lesions

VIII. Developmental or acquired deformities and conditions

Classification of periodontal diseases (AAP, 1999)

Most common:

- Chronic marginal gingivitis (CMG)

 - Inflammatory reaction to plaques

 - Reversible inflammation

- Chronic inflammatory periodontal disease (CIPD)

 - Adult periodontitis

 - Irreversible damage

 - Smoking important exacerbating factor

Pathophysiology

Bacteria

>600 species in the oral cavity
~200 detectable in an individual

8 bacterial species have been associated with periodontal disease
e.g.: *Prevotella intermedia* – acute necrotizing ulcerative gingivitis
Porphyromonas gingivalis – chronic inflammatory periodontal disease
Found in both healthy and diseased sites...
~ 50% of plaque bacteria can be cultured, rest are unknown!

Pathogenic factors:

- leukotoxins
- endotoxin
- capsular products (activators of bone resorption)
- hydrolytic enzymes (collagenases, phospholipases, proteases... etc)

Bacteria and bacterial toxins can invade the periodontal epithelium

TABLE 1 | The predominant human oral microbiota.

Microbial group	Microbial genus/species
Gram-positive	
Aerobic or facultative	<i>Streptococcus</i> (<i>S. gordonii</i> , <i>S. mitis</i> , <i>S. auralis</i> , <i>S. salivarius</i>) <i>Staphylococcus</i> (<i>S. aureus</i> , <i>S. epidermidis</i>) <i>Enterococcus</i> (<i>E. faecalis</i>) <i>Lactobacillus</i> (<i>L. casei</i> , <i>L. fermentum</i>) <i>Corynebacterium</i> (<i>C. matruchotii</i>) <i>Actinomyces</i> (<i>A. naeslundii</i> , <i>A. israeli</i> , <i>A. viscosus</i>) <i>Arachnia</i> (<i>A. propionica</i>) <i>Rothia</i> (<i>R. dentocariosa</i>)
Obligate anaerobes	<i>Bacillus</i> (<i>B. cereus</i>) <i>Propionibacterium</i> (<i>P. acnes</i>) <i>Peptostreptococcus</i> (<i>P. micros</i> , <i>P. anaerobius</i>)
Gram-negative	
Aerobic or facultative	<i>Campylobacter</i> (<i>C. rectus</i> , <i>C. concisus</i> , <i>C. gracilis</i>) <i>Actinobacillus</i> (<i>A. actinomycetemcomitans</i>)
Obligate anaerobes	<i>Fusobacterium</i> (<i>F. nucleatum</i>) <i>Porphyromonas</i> (<i>P. gingivalis</i>) <i>Prevotella</i> (<i>P. melaninogenica</i> , <i>P. oralis</i> , <i>P. intermedia</i>)

Pathophysiology

Immunogenetic factors

-*HLA association* (animal and human studies)

HLA-A9: associated with higher risk for CIPD, juvenile periodontitis, rapidly progressing periodontitis

indicate that HLA-A9 is associated with periodontal destruction

-*Genotype variants*

IL-1 α , IL-1 β , TNF α ; IL-4, IL-10

-*Twin studies*

No difference in gingivitis, probing depth, attachment loss, and plaque in monozygous twins raised apart or together

indicate that genetic component is more important than environment

-*Antibody response*

Usually directed against Gram- bacteria; levels correlate with disease severity

e.g. increased antibody levels against *P. gingivalis* in CIPD

Both systemic and local

Pathophysiology

Stages (*gingivitis always precedes periodontal disease!*)

I. Initial lesion: reversible damage to gingival sulcus, polymorphonuclear cell infiltration, complement activation

II. Early lesion: still reversible, lymphocytes replace polymorphonuclear cells. Mostly T cells, few plasma cells

III. Established lesion: predominant plasma cell infiltration, mainly IgG⁺

IV. Advanced lesion: destructive state; pocket formation, epithelial ulceration, periodontal ligament destruction, bone resorption

P. gingivalis important!

“PSD” model: polymicrobial synergy and dysbiosis

Cytokines

Cytokines & Periodontal Disease

BACTERIAL CHALLENGE

CYTOKINE RESPONSE

INNATE IMMUNITY

PMN



MØ



TNF- α , IL-1, IL-6

CHARACTERISTIC
CYTOKINES

PROTECTIVE
FEATURES

DESTRUCTIVE
FEATURES

No literature
evidence

Pro-inflammatory
RANKL inducers

ADAPTIVE IMMUNITY

Th1



IFN- γ

Anti-osteoclastogenic
IFN- γ *in vitro*

Pro-inflammatory
Th1 cells: RANKL+

Th17



IL-17

No literature evidences
Th1/Th2 inhibition (?)

Pro-inflammatory
Th7 cells: RANKL+
& RANKL inducers

Th2



IL-4

Anti-osteoclastogenic
IL-4 and IL-10 - *in vivo* & *in vitro*

B-cell lesion hypothesis
B cells: RANKL+

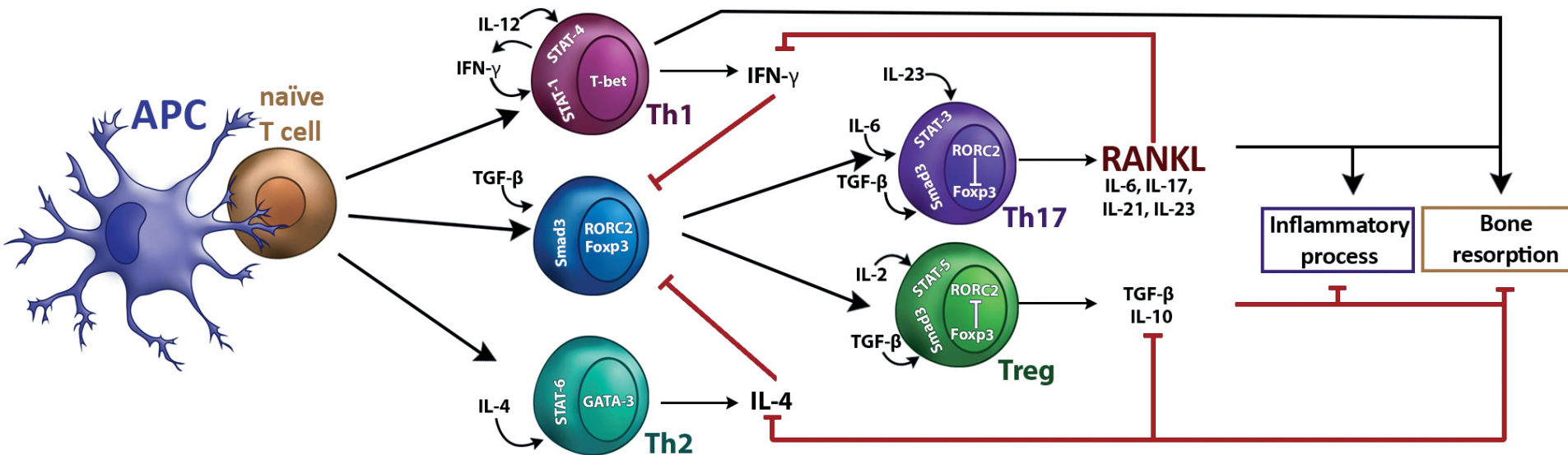
Tregs



IL-10, TGF- β

No literature
evidence

Osteoimmunology

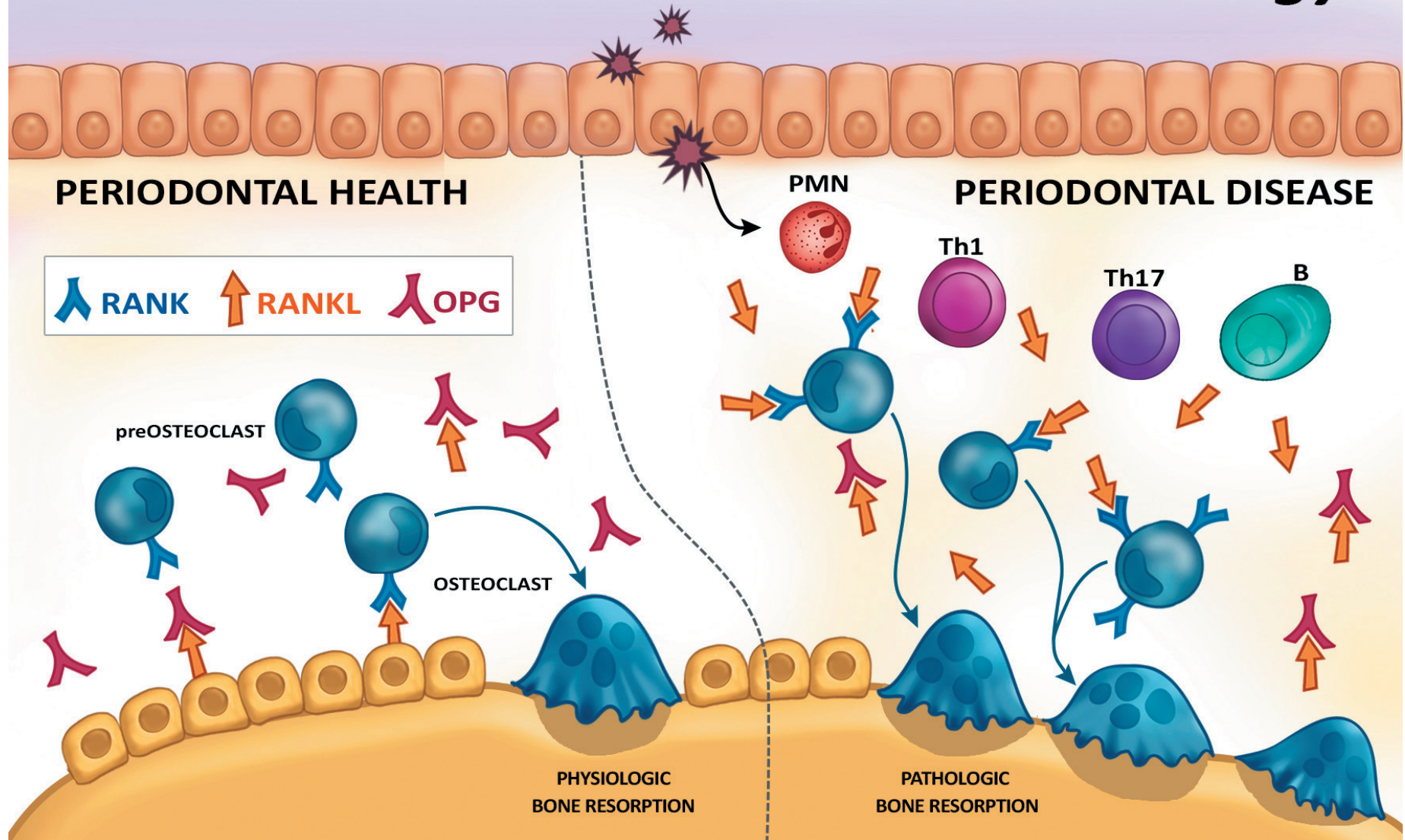


Osteoblast – Osteoclast balance:

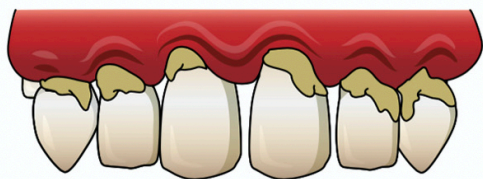
- RANKL: binds to RANK $\rightarrow$ Osteoclast differentiation, activation
- Osteoprotegerin: binds RANKL $\rightarrow$ inhibits osteoclast activation
- T_H17 cells can produce RANKL

Osteoimmunology

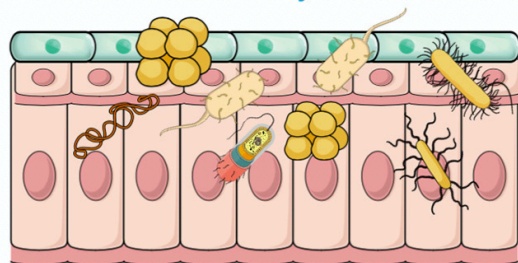
Periodontal Disease Osteoimmunology



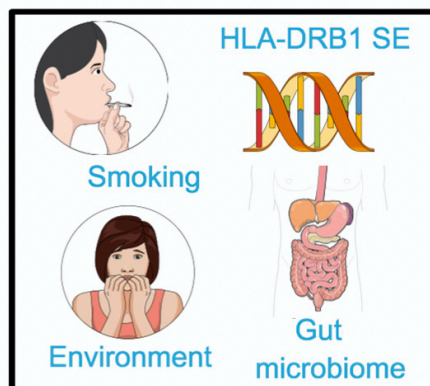
Periodontitis



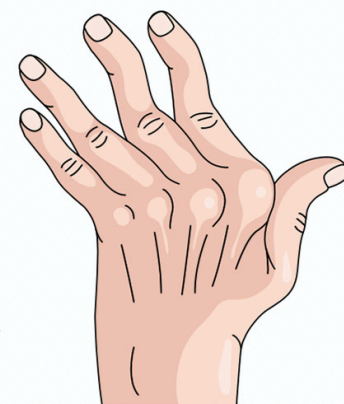
Microbial dysbiosis



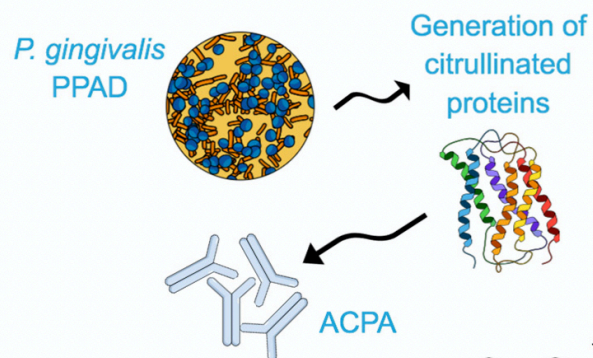
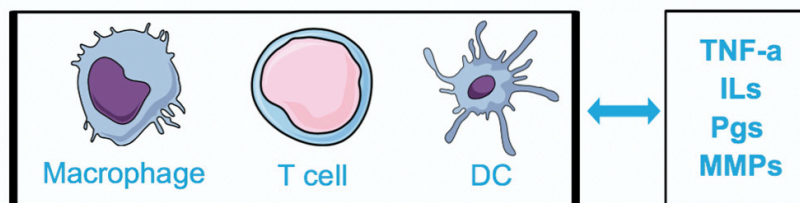
Risk factors



Arthritis



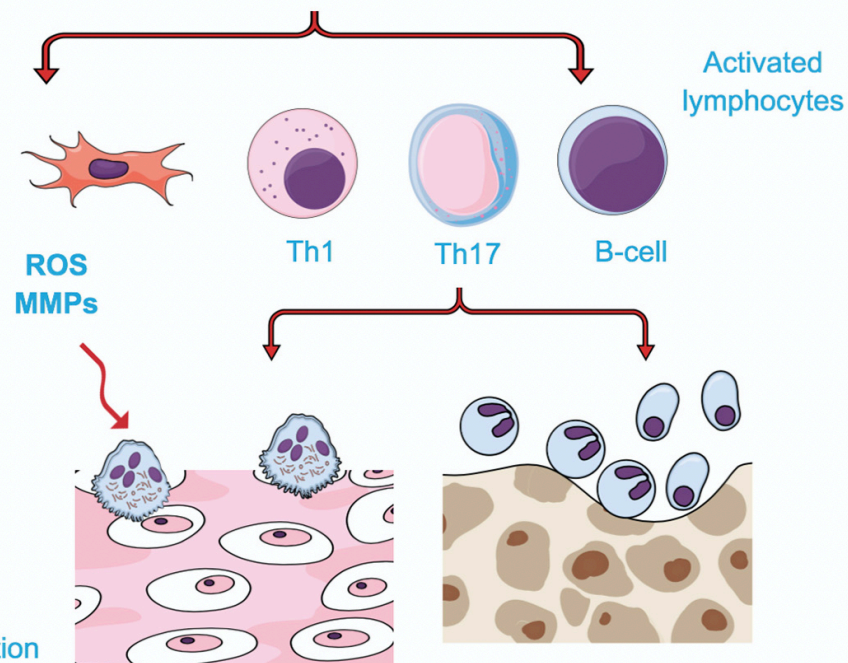
Development of
ACPA and RF



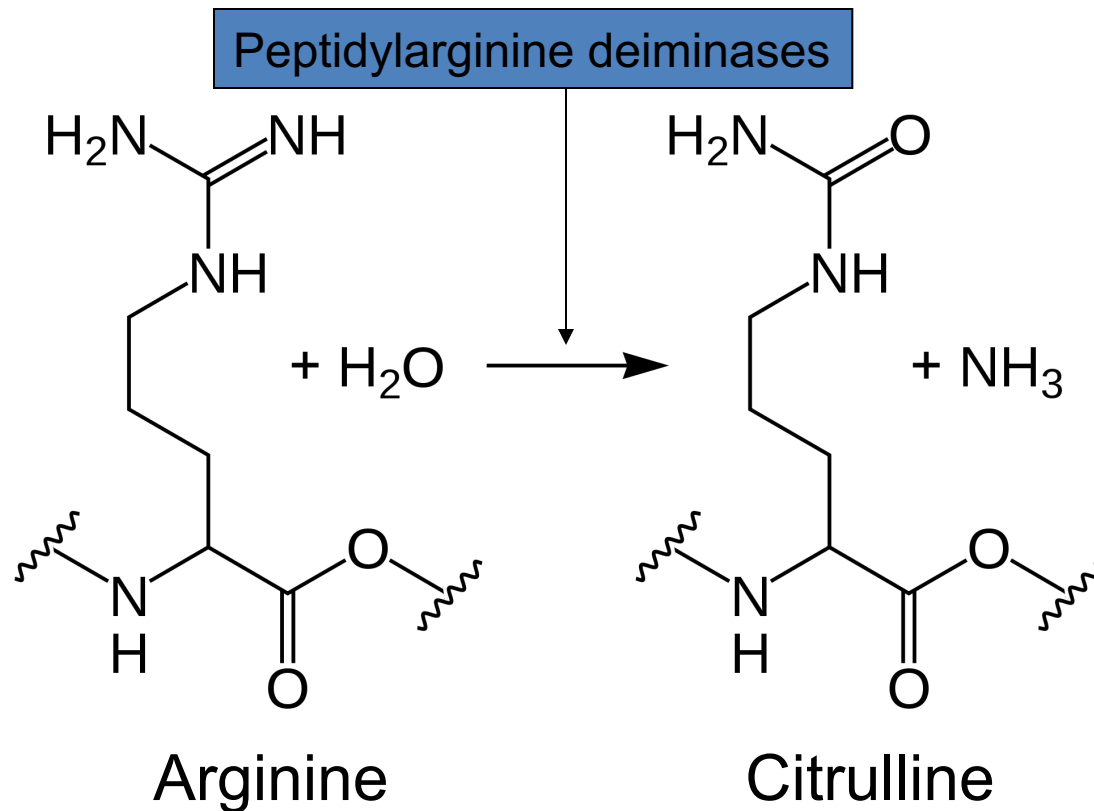
Formation of RF
Autoantibodies
Citrulline and
Antigen-presenting cell

Hypocitrullination in PMN
Activation of citrulline
enzymes

Bone and cartilage destruction



Citrullination (deimination) 1.



Citrullinated proteins:

MBP, filagrin, histon proteins, **vimentin**, fibrin, fibrinogen

MCV=modified citrullinated
vimentin

Intermediate filament family -
connective tissue, cytoskeleton

